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# Design, synthesis and cytotoxicity of cell death mechanism of rotundic acid derivatives

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# ABSTRACT

In the present investigation, 16 new rotundic acid (RA) derivatives modified at the C-3, C-23 and C-28 positions were synthesized. The cytotoxicities of the derivatives were evaluated against HeLa, A375, HepG2, SPC-A1 and NCI-H446 human tumor cell lines by MTT assay. Among these derivatives, compounds **4–7** exhibited stronger cell growth inhibitory than RA and compound **4** was found to be the best inhibition activity on five human tumor cell lines with  $IC_{50} < 10 \,\mu$ M. The apoptosis mechanism of compound **4** in HeLa cells was investigated by western blot analysis. The results indicated that compound **4** could induce apoptosis through increasing protein expression of cleaved caspase-3 and Bax, and decreasing protein expression of Bcl-2. In summary, the present work suggests that compound **4** might serve as an effective chemotherapeutic candidate.

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In the history of Traditional Chinese Medicine (TCM), medicinal plants and their extracts were used to treat various diseases. Nowadays, compounds derived from natural medicines with the unique and diverse chemical entities still constituted a considerable resource for developing novel medicaments. Over the recent years, a variety of biologically active constituents, including ginsenoside Rg3,<sup>1–3</sup> paclitaxel,<sup>4,5</sup> platycodin D,<sup>6</sup> triptolide,<sup>7</sup> and flavone eupatorin,<sup>8</sup> have been isolated from these sources and confirmed to have anti-cancer activity in both experimental and epidemiologic investigations. The bark of *Ilex rotunda* Thumb., was early recorded in 'Ling Nan Cai Yao Lu', has been used for a long time as a TCM for the treatment of colds, tonsillitis, pharyngitis, bone pain, acute gastroenteritis, and dysentery so on.<sup>9–11</sup> Nowadays, this medical plant is listed in Pharmacopoeia of the People's Republic of China, 2010.<sup>12</sup>

Rotundic acid  $(3\beta,19\alpha,23$ -trihydroxy-urs-12-en-28-oic acid, RA), a pentacyclic triterpene acid, is the major component isolated from the dry bark of *I. rotunda*. In addition to existence in aquifoliaceae plants,<sup>13–16</sup> RA also was isolated from *Mussaenda Pubescens*,<sup>17</sup> *Guettarda platypoda*,<sup>18</sup> *Olea europaea*,<sup>19</sup> *Planchonella* 

*duclitan*,<sup>20</sup> *Nauclea officinalis*.<sup>21</sup> Although RA could be obtained from the above plant resources, there were little reports on its bio-activity. To date, in addition to the anti-tumor<sup>20,22</sup> and lowering blood pressure activity,<sup>23</sup> no reports on other activities related to RA were published. Our studies have proved that RA had the activity of prevention and treatment of cardiovascular disease and we have applied two patents (one of the patent had been authorized).<sup>24,25</sup> More activities of RA need to be studied in the future.

In our previous study,<sup>26,27</sup> the results showed that a few of new amino acid derivatives showed stronger cytotoxicities than RA. In order to find the compounds with better cytotoxicities, we continued to carry out chemical modification and antitumor activity of RA. Since the nitrogen-containing organic compounds had high biological activities, which play an important role on the chemical research, so many researchers carried out research in this area.<sup>28</sup> In the work described herein, we focus on increasing the cytotoxicity of RA. As shown in Figure 1, due to higher steric hindrance, 19-OH is difficult to modified, Therefore, RA can be modified easily at C-3, C-23 and C-28 position. In the present study, we intend to modify RA to obtain better derivatives, their cytotoxicities are determined on the five human tumor cell lines including A-375 (human malignant melanoma cells), SPC-AI (human lung adenocarcinoma), Hela (cervical cancer cells), HepG2 (hepatoma cells) and NCI-H446 (small cell lung cancer). The derivatives with low IC<sub>50</sub> value will be further evaluated to explore its mechanism on the traditional apoptosis signal pathway.

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Figure 1. Structure of rotundic acid (RA).

Recently, the antitumor mechanism mainly focused on their effect on trigger apoptosis. Apoptosis,<sup>29</sup> known as programmed cell death, is closely related to many anticancer reagents. It has been broadly accepted that mitochondria play an important role during drug-induced apoptosis in cancer cells. Many of the stimulus including anticancer drug that triggered apoptosis on the mitochondria, which respond to proapoptotic signals by releasing cytochrome C, which belongs to an effective catalyst of apoptosis. Members of the Bcl-2 family include either proapoptotic (Bax) or antiapoptotic (Bcl-2) function, could lead to mitochondrial death signaling through cytochrome C release. Normally, Bcl-2 is local-



**Scheme 1.** Synthesis of RA derivatives. Reagents and conditions: (a) pyridine, acetic oxide, 80 °C, 16 h, 69.9%; (b) CH<sub>2</sub>Cl<sub>2</sub>, oxalyl chloride, rt, 20 h, 77.8–80.5%; (c) CH<sub>2</sub>Cl<sub>2</sub>, amino alcohols, rt, 1.5 h, 66.5–77.5%; (d) 1 mol/l NaOH in 60% methanol, 100 °C, 8 h, 88.4–90.1%; (e) CH<sub>2</sub>Cl<sub>2</sub>, acetic oxide, 4-DMAP, rt, 4 h, 79.5–90.1%; (f) CH<sub>2</sub>Cl<sub>2</sub>, Et<sub>3</sub>N, Ar-R<sup>2</sup>-NH<sub>2</sub>, rt, 1.5 h, 71.8–72.5%; (g)1 mol/l NaOH in 60% methanol, 100 °C, 6 h, 86.8–90.2%.

izes to the mitochondria and blocks the activation of proapoptotic proteins, such as Bax, to the mitochondria. When the cancer underwent chemotherapy, proapoptotic Bax can be upregulated in response to sensing DNA damage, Bax in turn stimulates mitochondria to release cytochrome C. Cytochrome C is the major inducer of caspase activation downstream of mitochondria. Finally, it will activate caspase-3 to be cleaved and execute apoptosis.

As reported in our previous investigation, RA was isolated and purified from *I. rotunda*,<sup>26</sup> its purity was  $\geq$  98% (HPLC assay) and the extraction yield of RA was much higher up to 100 mg/g. In this Letter, the synthetic routes of RA derivatives are outlined in Scheme 1. RA (1) was first converted to its 3,23-O-diacetate (2), which was then treated with oxalyl chloride to give the 28-acyl chloride (**3**).<sup>30</sup> This intermediate was then reacted with the appropriate amino alcohols (2-amino ethanol, 2-amino-1-propanol, 3amino-1-propanol, 2-amino-benzyl alcohol) in the presence of triethylamine to give the compounds N-[3B.23-diacetoxy-19\alpha-hydroxy-urs-12-en-28-oyl]-2-amino alcohols (compounds 4-7) Hydrolyzation of compounds 4-7 gave the corresponding N-[3<sub>β</sub>,19<sub>α</sub>,23-thrihydroxy-urs-12-en-28-oyl]-2-amino alcohols (compounds 8-11). Compounds N-[3 $\beta$ ,23-diacetoxy-19 $\alpha$ -hydroxy-urs-12-en-28-oyl]-2-amino alcohol acetates (compounds 12-**15**) were obtained by re-acetylating compounds **4**–7 in the presence of 4-dimethyl aminopyridine (4-DMAP), respectively. Compound 3 was treated with phenylamine or benzylamine in the presence of triethylamine to give the N-[3 $\beta$ ,23-diacetoxy-19 $\alpha$ -hydroxy-urs-12-en-28-oyl]-aromatic amine (compounds 16-17), hydrolyzation of compounds 16-17 to gave the corresponding N-[3β,19α,23-thrihydroxy-urs-12-en-28-oyl]-2-aromatic amine (18-19). The structures of these synthesized compounds were confirmed by Infrared (IR), Mass spectra (HR-ESI-MS), <sup>1</sup>H NMR and <sup>13</sup>C NMR assay.<sup>31–34</sup> The all 16 compounds obtained here were synthesized and reported for the first time, the purity of all these 16 compounds were above 98% with higher yield (HPLC).

In the present study, five kinds of human tumor cell lines including A-375, SPC-AI, Hela, HepG2 and NCI-H446 were the model chosen to determine the cytotoxicity of RA (as a positive control) and its derivatives **4–19**. Antiproliferative effects were determined with MTT assay.<sup>35</sup> Each experiment was repeated at least three times. The results are shown in Table 1.

As shown in Table 1, compound **2** (esterification of the 3-OH, 23-OH of RA) had stronger inhibitory activity against the five tu-

**Table 1** The  $IC_{50}$  values of RA and its derivatives on human cancer cell lines ( $\mu$ M)

mor cell lines with IC<sub>50</sub> values of 11.41, 23.24, 13.09, 6.85 and 7.11 µM, respectively. Significant further improvement of the cell growth inhibition on the five cell lines was achieved when the selected amino alcohols was coupled to the C-28 to yield compounds 4-7. However, compounds 8-11, hydrolyzation product of corresponding compounds 4-7, were found to exhibit lower anti-tumor effect than compounds 4-7, and the anti-tumor activity of compounds 8-11 were even weaker than it's lead compound RA. Though compounds 12–15 (esterification product of corresponding compounds 4-7) showed higher anti-tumor activity than compounds 8-11, they showed slightly lower anti-tumor activity compared to compounds **4–7**, the activity on A375 and Hela, even as same as compounds **4–7**. The results suggested that 3,23-O-diacetate group might be important to the inhibitory activity of tumor cell growth and free 3,23-OH might be detrimental to the inhibitory activity. In addition, from the above results we can speculate the reason that the activity of compounds **4–7** higher than **12–15** might be that compounds 4-7 has free hydroxyl group at the C-28 amide side chain. Because of the existence of free hydroxyl groups, together with the 3,23-O-diacetate group, may change ester water partition coefficient of compounds 4-7. As for compounds 16-19, the anti-tumor activity was not satisfactory as expected. Especially compounds 18-19, there was almost no inhibitory activity on the five tumor cell lines at all. However, the inhibitory activity of **16–17** was significantly higher than its hydrolysis product compounds 18-19. The result also gave the evidence that 3,23-O-diacetate group should be the essential group to increase the activity of RA.

From the Table 1, we can also notice that the anti-tumor activity of compound **4** is the highest among these compounds. The IC<sub>50</sub> value of compound **4** was significantly less than RA treatment group (<10  $\mu$ M) on the five cell lines with IC<sub>50</sub> values of 5.49, 3.61, 2.83, 4.40, and 6.67  $\mu$ M, respectively. These results indicated that compound **4** can be the candidate for the development of new anticancer drug. To explore its elementary anticancer effect, the apoptosis signal related protein level will be tested in the following experiment.

As seen in above experiment, result about the cytotoxicity of compound **4** was excited. The  $IC_{50}$  is much lower in compound **4** treatment for 24 h compared to RA group and other derivatives. In this experiment, we treated Hela cell with compound **4** at 10, 20 and 40  $\mu$ M for 24 h, Cisplatin (10  $\mu$ g/ml) was used as positive

Compound	IC <sub>50</sub>				
	A375	SPC-A1	Hela	HepG2	NCI-H446
1	16.58 ± 1.22	83.95 ± 3.28	31.92 ± 2.01	$7.33 \pm 0.68$	$11.40 \pm 2.32$
2	$11.41 \pm 0.99$	23.24 ± 1.26	$13.09 \pm 0.88$	$6.85 \pm 0.63^*$	$7.11 \pm 0.45$
4	$5.49 \pm 1.02^*$	$3.61 \pm 1.59^*$	$2.83 \pm 1.01^{\circ}$	$4.40 \pm 0.97^{*}$	$6.67 \pm 1.12^{*}$
5	$4.77 \pm 3.00^{*}$	12.88 ± 3.55	11.35 ± 3.26	$7.30 \pm 0.89^{*}$	$5.51 \pm 0.78^{\circ}$
6	$3.28 \pm 0.88^{*}$	13.36 ± 2.11	$6.97 \pm 1.97^{*}$	$5.49 \pm 1.23^*$	$5.08 \pm 1.26^{\circ}$
7	$3.71 \pm 0.75^*$	$8.09 \pm 1.46^{*}$	$3.32 \pm 0.48^{*}$	$4.45 \pm 1.03^{*}$	$7.03 \pm 0.92^{*}$
8	$21.43 \pm 2.01$	>100 <sup>a</sup>	>100 <sup>a</sup>	$45.60 \pm 2.03$	24.72 ± 1.87
9	35.48 ± 1.96	>100 <sup>a</sup>	$28.84 \pm 1.29$	$55.09 \pm 2.02$	39.81 ± 2.45
10	$12.47 \pm 2.01$	83.18 ± 1.89	17.74 ± 1.55	77.27 ± 2.13	$20.89 \pm 2.26$
11	18.97 ± 0.69	>100 <sup>a</sup>	$12.27 \pm 0.86$	>100 <sup>a</sup>	13.96 ± 1.12
12	$4.17 \pm 0.12^*$	$9.37 \pm 0.69^*$	$5.39 \pm 0.37^{*}$	$4.44 \pm 0.48^{*}$	18.62 ± 0.66
13	$5.10 \pm 0.89$	18.85 ± 1.77	$5.30 \pm 0.36$	$9.12 \pm 1.25$	14.69 ± 1.33
14	$10.06 \pm 1.61^*$	11.96 ± 0.93	$4.31 \pm 0.91^{*}$	11.96 ± 1.28	12.82 ± 1.83
15	$9.12 \pm 0.66^*$	13.21 ± 1.55	$4.27 \pm 2.33^{*}$	$12.35 \pm 2.78$	69.69 ± 2.01
16	$17.74 \pm 1.03$	$16.21 \pm 1.08$	$15.78 \pm 2.01$	$22.23 \pm 2.26$	$24.83 \pm 0.75$
17	13.71 ± 1.00	$12.10 \pm 2.06$	$19.18 \pm 2.23$	$19.50 \pm 0.99$	$26.92 \pm 1.09$
18	70.79 ± 0.85	>100 <sup>a</sup>	$23.17 \pm 0.76$	$64.65 \pm 0.69$	$70.79 \pm 0.78$
19	>100 <sup>a</sup>	>100 <sup>a</sup>	$28.05 \pm 1.12$	>100 <sup>a</sup>	>100 <sup>a</sup>

Notes: Data are represented in mean  $\pm$  SD; n = 3.

 $^a$  IC\_{50} values more than 100  $\mu M$  are indicated as >100.

 $^*$  P < 0.05 versus RA and IC<sub>50</sub> value < 10  $\mu$ M.



**Figure 2.** Alteration of protein expression of Bax, Bcl-2, cytochrome C in Hela cells after 24 h compound **4** treatment. (A) Protein expression of Bcl-2; (B) protein expression of Bax; (C) the ratio of Bax to Bcl-2; (D) protein expression of cytochrome C; (E) presented compound **4**, C presented cisplatin, \**P* <0.05 compared to control group, \*\**P* <0.01 compared to control group.

treatment group. The protein expression of Bax, Bcl-2, cytochrome C and caspase-3 were detected by western blot. As shown in Figure 2, the antiapoptosis effect of compound **4** presented dose dependent manners after 24 h treatment, Compound **4** could decrease the protein expression of Bcl-2, a antiapoptotic member of Bcl family, and increase a proapoptotic protein Bax level, the ratio of Bax to Bcl-2 was remarkable increased in a dose dependent manner. The key protein of mithochondial related apoptosis signal, cytochrome C in the cytoplasm also dramatically increased follow-

ing the compound **4** treatment. All these result indicated that compound **4** might block the expression of Bcl-2, which could localize to the mitochondria and block the activation of proapoptotic proteins, such as Bax, to the mitochondria. The ratio of Bax to Bcl-2 has been used in many studies as the evaluation the level of apoptosis. The present study, compound **4** increased this ration by elevated Bax expression and depressed Bcl-2 expression finaly trigger the cytochrome C release from mitochondrial. It has been broadly accepted that the alteration on Bcl-2, Bax was closely related to



**Figure 3.** Alteration of protein expression of caspase-3 and cleaved caspase-3 in Hela cells after 24 h compound **4** treatment. (A) Protein expression of caspase-3; (B) protein expression of cleaved caspase-3, E presented compound **4**; (C) presented cisplatin, \**P* <0.05 compared to control grup, \*\**P* <0.01 compared to control group.

stimulation of apoptosis, the effector of their alteration presented as the cytochrome C releasing, which will trigger the cleave of executor caspase, caspase-3. As shown in Figure 3, the protein expression of caspase-3 was significantly elevated by compound 4 treatment, the cleaved protein expression of caspase-3 remarkabley increased as well. It has been broadly reported that caspases are widely expressed in an inactive proenzyme form in most cells and once activated can often activate other procaspases. Caspase-3 among the caspase family is considered to be the most important of the executioner caspases. Caspases-3 is cysteine proteases, using the sulfur atom in cysteine to cleave polypeptide chains. And the cleaved capase-3 is the active form when the apoptosis was triggered. The cleaved caspase-3 will finally induce the DNA fragmentation in cancer cell. Therefore, the elevation of cleaved caspase-3 is very important for the compound which affects cancer growth by increasing apoptosis. Our result showed that compound **4** could increase the protein level of cleaved caspase-3 in a dose dependent

cal effect by promoting apoptosis in Hela cells. In conclusion, we have synthesized sixteen RA derivatives and tested their anti-tumor activity in vitro for the first time. The result showed that compounds 4-7 have stronger inhibitory activity on tumor cell growth than RA. Among these RA derivatives, compound 4 was found with the best inhibition activity on the five human tumor cell lines with  $IC_{50}$  less than 10  $\mu$ M. From the results, we can conclude that 3,23-O-diacetate group might be important to the inhibitory activity of tumor cell growth and free 3,23-OH might be detrimental to the inhibitory activity. In view of the best activity of compound 4, we further carried out the investigation on the anticancer mechanism on Hela cells. The results indicated that compound **4** could suppress the growth of Hela human cells, and induce apoptosis by increasing the protein expression of cleaved caspase-3and Bax, and decreasing the protein expression of Bcl-2. Therefore, compound **4** may serve as a potential lead compounds for new anticancer drug development.

manner. Taken together, compound 4 has an active pharmacologi-

In summary, our study provided a powerful incentive for further research on the chemical modifications, and laid the foundation for further study of RA derivatives and their structureactivity relationships. Moreover, our study also provided theoretical basis on finding safe and effective anti cancer lead compounds from the natural medicine.

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## Supplementary data

Supplementary data associated with this article can be found, in the online version, at http://dx.doi.org/10.1016/j.bmcl.2013. 03.005. These data include MOL files and InChiKeys of the most important compounds described in this article.

#### **References and notes**

- Lee, C. K.; Park, K. K.; Chung, A. S.; Chung, W. Y. Food Chem. Toxicol. 2012, 50, 2565.
- Jiang, J. W.; Chen, X. M.; Chen, X. H.; Zheng, S. S. World J. Gastroenterol. 2011, 17, 3605.
- 3. Guo, G.; Xu, J. H.; Sun, J.; Fan, Z. Z. World J. Gastroenterol. 2012, 20, 1004.
- Xu, Q.; Liu, Y. X.; Su, S. S.; Li, W.; Chen, C. Y.; Wu, Y. Biomaterials 2012, 33, 1627.
  Zhao, B. X.; Huang, Y.; Luo, L. M.; Zhao, X.; Wang, X.; Chen, S.; Yu, K. F.; Zh, X.; Zh, Q. J. Pharm. Sci. 2012, 21, 40.
- 6. Yu, J. S.; Kim, A. K. J. Med. Food **2010**, 13, 298.
- Huang, W. W.; He, T. T.; Chai, C. S.; Yang, Y.; Zheng, Y. H.; Zhou, P.; Qiao, X. X.; Zhang, B. PLoS ONE 2012, 7, e37693.
- Ková, I. D.; Rárová, L.; Grúz, J.; Vondrusová, M.; Strnad, M.; Kryštof, V. Fitoterapia 2012. FITOTE-02465.
- 9. Huang, W. J. Jiangxi J. Tradit. Chin. Med. 1996, 2, 147.
- 10. Wen, D. X.; Chen, Z. L. Phytochemistry 1995, 41, 657.
- The Chinese Ministry of Public Health, the Institute of Drug Control. The Colour Atlas of Chinese Herbs. 1987, 245.
- 12. Pharmacopoeia of the People's Republic of China, 2010, 293.
- Sun, H.; Zhang, X. Q.; Cai, Y.; Han, W. L.; Wang, Y.; Ye, W. C. Chem. Ind. Forest Prod. 2009, 29, 111.
- 14. Xie, J. B.; Bi, Z. M.; Li, P. Acta Pharm. Sin. 2003, 38, 534.
- Liao, L. P.; Bi, Z. M.; Li, P.; Xie, J. B.; Zhang, T. D. Chin. J. Nat. Med. 2005, 3, 344.
  Haraguchi, H.; Kataoka, S.; Okamoto, S.; Hanafi, M.; Shibata, K. Phytother. Res.
- **1999**, *13*, 151. 17. Zhao W. M. Wolfender, I.L. Hostettmann, K. Cheng, K. F. Xu, R. S. Oin, G. W.
- 17. Zhao, W. M.; Wolfender, J. L.; Hostettmann, K.; Cheng, K. F.; Xu, R. S.; Qin, G. W. *Phytochemistry* **1997**, *45*, 1073.
- 18. Bhattacharyya, J.; Almeida, M. D. J. Nat. Prod. 1985, 48, 148.
- Saimaru, H.; Orihara, Y.; Tahsakul, P.; Kang, Y. H.; Shibuya, M.; Ebizuka, Y. Chem. Pharm. Bull. 2007, 55, 784.
- 20. Li, Z. H.; Zhuang, S. H.; Xu, F. L.; Wu, Z. Y. J. Chin. Chem. Soc. 2005, 52, 1275.
- 21. Ma, W. Z.; Ling, T. J.; Zhang, Y. H.; Lin, L. D. J. Trop. Subtrop. Bot. 2005, 13, 167.
- 22. Xu, R. Guangzhou University of Chinese Medicine. 2009.
- 23. Liu, G. Q. CHN 200810198190.7, 2010.
- 24. Zhao, Q. C.; Nan, M. L.; He, Y. F.; Chen, S. W. CHN ZL201010204596.9, 2010.
- Zhao, Q. C.; He, Y. F.; Nan, M. L.; Chen, S. W.; Zhao, Y. W.; Wang, L. P. CHN 20110030007.4, 2011.
- He, Y. F.; Nan, M. L.; Sun, J. M.; Meng, Z. J.; Yue, F. G.; Zhao, Q. C.; Yang, X. H.; Wang, H. Molecules 2012, 17, 1278.
- Chen, Y.; He, Y. F.; Nan, M. L.; Sun, W. Y.; Hu, J.; Cui, A.; Li, F.; Wang, F. Int. J. Mol. Med. 2013, 31, 353.
- 28. Xin, G. Y. CHN: Southeast University. 2007.
- 29. Fulda, S.; Debatin, K. M. Cancer Lett. 2003, 197, 131.
- 30. Meng, Y. Q.; Liu, D.; Zhao, J.; Wang, Z. CHN: 200610047236.6, 2006.
- Kim, N. C.; Desjardins, A. E.; Wu, C. D.; Kinghorn, A. D. J. Nat. Prod. 1999, 62, 1379.
- 32. Nakatani, M.; Hatanaka, S.; Komura, H.; Kubota, T.; Hase, T. *Chem. Soc. Jpn.* **1989**, *62*, 469.
- 33. Zhang, A. L.; Ye, Q.; Li, B. G.; Qi, H. Y.; Zhang, G. L. J. Nat. Prod. 2005, 68, 1531.
- 34. Xie, G. B.; Zhou, S. X.; Lu, Y. N.; Lei, L. D.; Tu, P. F. Chem. Pharm. Bull. 2009, 57, 520.
- 35. Mosmann, T. J. Immunol. Methods 1983, 139, 55.